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Serum surfactant protein A as a surrogate biomarker of a negative heart sign among patients with interstitial lung disease

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ABSTRACT

The mechanisms underlying interstitial lung disease (ILD) are characterized by variable inflammation or fibrosis of the pulmonary interstitium. A negative heart sign (NHS) on ⁶⁷Ga scintigrams of patients with ILD is due to considerably increased inflammatory activity in the lungs. We retrospectively analyzed relationships between NHS and established biomarkers of disease severity in patients with ILD. Among 81 consecutive non-smoking patients with ILD (mean age, 63 years) who had been hospitalized between April 2009 and October 2011, we selected 52 who had been assessed by 67Ga scintigraphy. We then evaluated relationships between NHS and blood biomarkers, pulmonary function and high-resolution computed tomography (HRCT). Among these 52 patients, 10 showed idiopathic pulmonary fibrosis and 42 had other ILD. Multivariate analysis with stepwise variable selection, serum surfactant protein (SP)-A (OR (odds ratio), 1.026; 95%CI (confidence interval), 1.003-1.050; P = 0.024) and inflammation index calculated from HRCT findings (OR, 1.358; 95%CI, 1.079–1.709; P = 0.009) were significant predictors of an NHS. Serum SP-A offered 85% sensitivity and 75% specificity for predicting NHS at an optimal cut-off of 45.8 ng/ mL. Serum SP-A concentrations correlated positively with inflammation index (r = 0.344, P = 0.015). In conclusion, serum SP-A might serve as a surrogate biomarker for predicting an NHS in patients with ILD.

Keywords: inflammation, idiopathic pulmonary fibrosis, surfactant protein, treatment, gallium uptake

Abbreviations: %DLco: percentage predicted diffusion capacity of carbon monoxide %FVC: percentage predicted forced vital capacity %VC: percentage predicted vital capacity AE: acute exacerbation AUC: area under the receiver operating characteristics curve CI: confidence interval CTD-ILD: connective tissue disease-associated ILD ELD: eosinophilic lung disease HP: hypersensitivity pneumonitis HRCT: high-resolution computed tomography

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IgG4-RD: immunoglobulin G4-related disease ILD: interstitial lung disease iNSIP: idiopathic nonspecific interstitial pneumonia IPF: idiopathic pulmonary fibrosis NHS: negative heart sign OP: organizing pneumonia OR: odds ratio ROC: receiver operating characteristics SD: standard deviation SP: surfactant protein

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INTRODUCTION

The pathological mechanisms underlying interstitial lung disease (ILD) are characterized by various inflammatory processes or fibrosis of the pulmonary interstitium.¹ Evaluation of the severity of lung inflammation in patients with ILD is crucial when predicting disease activity and deciding therapeutic strategies. Various parameters for detecting lung inflammation (alveolitis) have been reported, including blood biomarkers (surfactant protein (SP)-A and SP-D^{2,3}), and high-resolution CT (HRCT) scores (inflammation index⁴). However, parameters allowing accurate evaluation of lung inflammation in clinical practice have yet to be established.

Increased uptake of ⁶⁷Ga on pulmonary scintigrams offers a reliable indicator of strong lung inflammation, with such uptake appearing brighter than cardiac (blood flow) images.⁵⁻⁷ This is referred to as a negative heart sign (NHS) (Figure 1).⁸ However, ⁶⁷Ga scintigraphy is unsuitable for rapid and simple assessments of lung inflammation in terms of labor and cost.

The present study retrospectively investigated data from patients who had been assessed using



Fig. 1 Representative images for presence and absence of a negative heart sign (NHS) Fig. 1A: Presence of an NHS. Fig. 1B: Absence of an NHS.

⁶⁷Ga scintigraphy, with the aim of identifying parameters available in clinical practice that can help to predict an NHS.

MATERIALS AND METHODS

Study location and patients

This study proceeded at the National Defense Medical College Hospital in Japan. We selected 52 non-smoking patients with ILD who had been admitted to hospital and assessed by ⁶⁷Ga scintigraphy between April 2009 and October 2011. Because serum SP-A is reportedly increased due to the effects of smoking, current smokers were excluded from this research.⁹ Patients were then assigned to groups depending on the presence or absence of an NHS on ⁶⁷Ga scintigraphy (Figure 2). Medical history, physical findings, blood biomarkers, pulmonary function and HRCT findings were compared between groups.

Diagnosis of ILD

Idiopathic pulmonary fibrosis (IPF) was diagnosed based on the IPF consensus classification.¹⁰ Patients without IPF were subdivided according to whether ILD was stable at the time of evaluation or exacerbated non-IPF was present, defined as acute, progressive disease requiring steroid pulse therapy and accompanied by fever, dry cough, and/or dyspnea.¹¹ Connective tissue disease-associated ILD (CT-ILD) was diagnosed in patients without IPF based on physical, serological and HRCT findings that were consistent with ILD. Lung biopsy specimens were histologically



Fig. 2 Flowchart of patient recruitment and analysis

evaluated to exclude other specific diseases. Idiopathic nonspecific interstitial pneumonia (iNSIP), organizing pneumonia (OP), eosinophilic lung disease (ELD), hypersensitivity pneumonitis (HP), pulmonary sarcoidosis and immunoglobulin G4-related disease (IgG4-RD) were diagnosed based on established criteria.^{10,12-16}

Pulmonary function tests and blood biomarkers

Blood samples were collected at the time of admission from all patients, and serum SP-A (normal < 43.8 ng/mL), SP-D (normal < 110 ng/mL) and KL-6 (normal < 500 U/mL) were measured. Lung function was tested within 1 month before admission and percentage predicted forced vital capacity (%FVC), percentage predicted vital capacity (%VC), and percentage predicted diffusion capacity of carbon monoxide (%DLco) were determined.

HRCT and 67Ga scintigraphy

Patients were evaluated by HRCT and ⁶⁷Ga scintigraphy within 1 month before admission and images were independently assessed by two pulmonologists and one radiologist. Findings from HRCT were evaluated using the semi-quantitative scoring method described by Ooi et al.⁴ Abnormalities on HRCT of the lungs were categorized as ground glass opacity, mixed ground glass and reticular disease, or reticular fibrosis and honeycomb lung, then scored based on ratios (%) of disease in each of the six lung lobes (Figure 3). Global scores were calculated by adding the scores for each anomaly in all lobes. An inflammation index was derived from the sum of scores for ground glass opacity, mixed ground glass and reticular disease. Fibrosis index was



Fig. 3 Abnormalities on HRCT of the lungs

Fig. 3A: Ground glass opacity.

- Fig. 3B: Mixed ground glass and reticular disease.
- Fig. 3C: Reticular fibrosis.
- Fig. 3D: Honeycomb lung.

calculated as the sum of reticular fibrosis and honeycomb scores.

Statistical analysis

Data were statistically analyzed using JMP version 11 software (SAS Institute, Cary, NC) and are expressed as mean \pm standard deviation (SD). Groups were compared using Wilcoxon ranksum tests. Optimal parameter cut-off values were determined from receiver operator characteristics (ROC) curves. Primary predictors for NHS were determined using multiple stepwise regression analysis. Nonparametric Spearman's rank correlation coefficients were calculated to assess correlations between SP-A and other clinical parameters. Values of P < 0.05 were considered significant.

Ethics approval

The institutional review board at the National Defense Medical College approved this study (approval number: kan-75; approval date: 19 October 2011). In all patients, consent for participation of this retrospective study was obtained by disclosing a clinical study including the description of opt-out (http://www.ndmc.ac.jp/wp-content/uploads/2016/03/test_state75.pdf).

RESULTS

Patient characteristics

Among 81 consecutive non-smoking patients with ILD, 52 were assessed by ⁶⁷Ga scintigraphy within 1 month before hospitalization. Among the remainder, 15 patients with severe respiratory failure who did not undergo scintigraphy and 14 patients who did not undergo scintigraphy within 1 month before hospital admission were excluded. Ten of the 52 patients were diagnosed with IPF. The remaining 42 patients had other types of ILD, comprising CT-ILD (n = 3; including polymyositis/dermatomyositis, rheumatoid arthritis and autoimmune hepatitis (n = 1 each)), iNSIP (n = 2), OP (unknown etiology n = 5; drug-induced, n = 1), ELD (eosinophilic granulomatosis with polyangiitis, n = 1; eosinophilic pneumonia n = 1), HP (n = 6), pulmonary sarcoidosis (n = 19) and IgG4-RD (n = 4). ILD was stable in 37 patients, with the remaining 5 patients showing exacerbated ILD (CT-ILD, n= 2; OP, n = 1; ELF, n = 2) at the time of evaluation.

Comparison of patients with and without NHS

Mean age of the 52 patients was 63 years, with 31 males (62%) and 48 patients (92%) showing pathologically confirmed ILD. Patients were categorized from ⁶⁷Ga scintigraphy as showing presence (n = 27) or absence (n = 25) of an NHS (Figure 2). Table 1 shows the characteristics of patients. Age, sex, smoking status, pathologically confirmed disease, diagnostic details and outcomes did not differ significantly between patients with and without an NHS. In contrast, serum SP-A, SP-D, KL-6, %VC, %FVC, %DLco, inflammation index and fibrosis index differed significantly between groups.

Predicting an NHS using ROC curves

Areas under ROC curves (AUCs) for serum SP-A, SP-D, KL-6, %VC, %FVC, %DLco, inflammation index and fibrosis index to predict an NHS were 0.83, 0.75, 0.78, 0.73, 0.71, 0.77, 0.75 and 0.67, respectively (Table 2). The AUC of all variables showed that serum SP-A could predict the presence of an NHS the most accurately among the measured parameters. At an optimal cut-off 45.8 ng/mL, serum SP-A offered 85% sensitivity and 75% specificity for predicting an NHS.

Characteristics	All patients	^a NHS +	NHS –	P NHS+ vs. NHS–
Total (n)	52	27	25	
Age (y)	63±12	64.9±9.4	60.0±14.2	0.193
Male sex, n (%)	31 (62)	18 (67)	13 (52)	0.282
Smoking status (Former / never)	30 / 22	18 / 9	12 / 13	0.173
Pathological proven disease, n (%)	48 (92)	24 (89)	24 (96)	0.336
Parameters				
Serum surfactant protein-A (ng/mL)	66.4±43.5	78.2±43.9	44.0±30.5	< 0.001
Serum surfactant protein-D (ng/mL)	212.2±223.8	287±272	131±116	0.003
Serum KL-6 (U/mL)	1182.8±1080.2	1565±1120	753±868	< 0.001
Vital capacity (%predicted)	95.2±24.1	86.0±24.4	105.2±19.7	0.007
Forced vital capacity (% predicted)	93.8±25.3	84.6±26.4	103.8±20.0	0.013
Diffusion capacity of carbon monoxide (%predicted)	73.9±23.0	64.8±24.9	83.3±16.7	0.001
Inflammation index	4.4±4.2	6.1±3.9	2.5±3.8	0.001
Fibrosis index	0.9±3.1	2.3±4.3	0.2±1.0	0.004
Diagnosis n (%)				
Idiopathic pulmonary fibrosis	10 (19)	6 (22)	4 (16)	0.570
Other interstitial lung diseases	42 (81)	21 (78)	21 (84)	0.570
Outcome				
Follow-up, days	707±402	667±396	749±413	0.404
Death, n (%)	5 (10)	2 (7)	3 (12)	0.575

 Table 1.
 Patient's characteristics

Footnote:

^aNHS: negative heart sign

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Variable	AUC	Best cut-offs	Sensitivity	Specificity	Р		
Serum surfactant protein-A (ng/mL)	0.83	45.8	85	75	0.002		
Serum surfactant protein-D (ng/mL)	0.75	184	74	75	0.011		
Serum KL-6 (U/mL)	0.78	603	81	67	0.015		
Vital capacity (%predicted)	0.73	80.9	56	87	0.009		
Forced vital capacity (%predicted)	0.71	90.4	68	78	0.012		
Diffusion capacity of carbon monoxide (%predicted)	0.77	68.5	75	87	0.010		
Inflammation index	0.75	7	59	88	0.003		
Fibrosis index	0.67	1	37	96	0.086		

Table 2. Analysis of ROC curves to predict negative heart sign.

Table 3. Multiple stepwise regression analysis of primary predictor of negative heart sign.

Variable	Odds ratio	95% Confidence interval	Р
Surfactant protein-A (ng/mL)	1.026	1.003-1.050	0.024
Inflammation index	1.358	1.079-1.709	0.009
Fibrosis index	1.433	0.903-2.274	0.127



Fig. 4 Relationship between serum SP-A concentrations and HRCT scores Serum SP-A concentrations correlate significantly with inflammation index (r = 0.344, P = 0.015), but not with fibrosis index (r = 0.103, P = 0.477).

Stepwise multivariate analysis

The variables of age, sex, smoking status, presence of pathologically confirmed disease, IPF or other ILD, serum SP-A, SP-D, KL-6, %VC, %FVC, %DLco, inflammation index and fibrosis index were assessed using stepwise multiple logistic regression. Serum SP-A (odds ratio (OR), 1.026; 95%CI, 1.003–1.050; P = 0.024) and the inflammation index calculated from HRCT findings (OR, 1.358; 95%CI, 1.079–1.709; P = 0.009) were significant predictors of an NHS (Table 3).

Relationship between serum SP-A and HRCT scores

Among patients with and without an NHS, inflammation indices were $6.1 \pm 3.9\%$ and $2.5 \pm 3.8\%$, respectively, while fibrosis indices were $2.3 \pm 4.3\%$ and $0.2 \pm 1.0\%$, respectively. Serum SP-A concentrations correlated with inflammation index (r = 0.344, P = 0.015), but not with fibrosis index (r = 0.103, P = 0.477) (Figure 4).

DISCUSSION

We aimed to determine an accurate, cost-effective and simpler means of detecting the severity and extent of lung inflammation in patients with ILD.

Lung inflammation due to the accumulation of lymphocytes, neutrophils, or eosinophils influences the pathogenesis of ILD. As a result, assessing lung inflammation in patients with ILD is thus important for evaluating disease activity.¹⁷⁻²⁰ Actually, 27 (52%) of 52 patients with an NHS as determined from ⁶⁷Ga scintigraphy in the present study displayed strong lung inflammation. Values for blood biomarkers (serum SP-A, SP-D, and KL-6), pulmonary function tests (%VC, %FVC, %DLco), and HRCT findings (inflammation and fibrosis indices) were worse among patients with, than without an NHS. Also, 4 of 5 patients (80%) with exacerbated non-IPF lung diseases requiring steroid pulse therapy had an NHS. We therefore speculated that in ILD patients with NHS, attention should be paid to the presence of strong lung inflammation, not only as an AE of ILD, but also in NSIP, OP, and ELD as pathologies for which the standard treatment is steroid administration. However, ⁶⁷Ga scintigraphy is not an appropriate modality in terms of radiation exposure and labor for patients in the clinical setting who have such strong lung inflammation that requires immediate and intensive treatment. Persistent lung inflammation due to oxidative stress worsens the prognosis for patients with ILD and accumulating evidence is showing that supportive therapy with anti-inflammatory macrolides and steroids (in addition to antifibrotic agents) is useful.²¹⁻²⁴ However, the present study did not investigate relationships between anti-inflammatory therapy according to the presence or absence of an NHS and disease prognosis.

Serum SP-A as a surrogate biomarker of lung inflammation in patients with ILD might be able to replace ⁶⁷Ga scintigraphy, which can accurately detect lung inflammation, but is too laborious and expensive for the clinical setting.⁵⁻⁷ SP-A is a member of the collectin family, and alveolar epithelial type II pneumocytes comprise the major source of surfactant apoproteins. Alveolar epithelial type II pneumocytes in lungs with alveolitis secrete SP-A, which is detectable in serum.^{1,2} Furthermore, concentrations of SP-A correlate with the extent of alveolitis (confirmed as HRCT findings of ground glass opacity), but not with progression of fibrosis.³ Consistent with this, we found that serum SP-A correlated with the inflammation index, which represents a robust indicator of alveolitis.⁴ As a prospective study of lung biopsy-confirmed IPF has associated elevated serum SP-A with risk of mortality,²⁵ whether baseline serum SP-A correlates with long-term prognosis in patients with ILD warrants investigation.

The previous and present findings indicate that serum SP-A could provide a cost-effective, simple and rapid alternative to ⁶⁷Ga scintigraphy. However, this single-institution study of a small number of patients shows some clear limitations that need to be kept in mind when interpreting our results. The repeatability of our findings requires evaluation in a multi-center prospective study. Clinical diagnoses of the enrolled patients were heterogenous, and the clinical relevance of serum SP-A values should therefore be evaluated for specific histopathological diagnoses (for example, IPF alone), although ILD subtypes were not identified as significant predictors of an NHS in the present study. In addition, evaluating the relationship between the cellularity in bronchoalveolar lavage fluid and serum SP-A may identify subtypes of inflammatory cells (neutrophilic or lymphocytic) that more closely correlate with increases in serum SP-A concentrations.

CONCLUSION

Serum SP-A might serve as a surrogate biomarker for predicting an NHS in patients with ILD.

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DISCLOSURE

None of the authors have any real or perceived conflicts of interest to declare regarding the subject of this manuscript.

REFERENCES

- Takahashi H, Shiratori M, Kanai A, et al. Monitoring markers of disease activity for interstitial lung diseases with serum surfactant proteins A and D. *Respirology*. 2006;11(suppl):S51–S54. doi: 10.1111/j.1440-1843.2006.00809.x.
- Ohnishi H, Yokoyama A, Kondo K, et al. Comparative study of KL-6, surfactant protein-A, surfactant protein-D, and monocyte chemoattractant protein-1 as serum markers for interstitial lung diseases. *Am J Respir Crit Care Med.* 2002;165(3):378–381. doi: 10.1164/ajrccm.165.3.2107134.
- 3. Takahashi H, Fujishima T, Koba H, et al. Serum surfactant proteins A and D as prognostic factors in idiopathic pulmonary fibrosis and their relationship to disease extent. Am J Respir Crit Care Med. 2000;162(3):1109–1114. doi: 10.1164/ajrccm.162.3.9910080.
- 4. Ooi GC, Mok MY, Tsang KW, et al. Interstitial lung disease in systemic sclerosis. *Acta Radiol*. 2003;44(3):258–264. doi: 10.1034/j.1600-0455.2003.00058.x.
- Line BR, Hunninghake GW, Keogh BA, et al. Gallium-67 scanning to stage the alveolitis of sarcoidosis: correlation with clinical studies, pulmonary function studies, and bronchoalveolar lavage. Am Rev Respir Dis. 1981;123(4):440–446. doi: 10.1164/arrd.1981.123.4.440.
- Crystal RG, Bitterman PB, Rennard SI, et al. Interstitial lung diseases of unknown cause. Disorders characterized by chronic inflammation of the lower respiratory tract (first of two parts). N Engl J Med. 1984;310(3):154–166. doi: 10.1056/NEJM198401193100304.
- 7. Liu FY, Shiau YC, Kao A, et al. Comparison of quantitative 99mTc-HMPAO and 67Ga citrate lung scans in patients with active diffuse infiltrative lung disease. *Nucl Med Commun.* 2003;24(12):1243–1246.
- Cooke SG, Davies ER, Goddard PR. Pulmonary uptake in 67-gallium citrate scintigraphy-the 'negative heart' sign. *Postgrad Med J.* 1989;65(770):885–891. Doi: 10.1136/pgmj.65.770.885.
- Kobayashi H, Kanoh S, Motoyoshi K. Serum surfactant protein-A, but not surfactant protein-D or KL-6, can predict preclinical lung damage induced by smoking. *Biomarkers*. 2008;13(4):385–392. doi: 10.1080/13547500801903651.
- Travis WD, Costabel U, Hansell DM, et al. An official American Thoracic Society/European Respiratory Society statement: update of the international multidisciplinary classification of the idiopathic interstitial pneumonias. Am J Respir Crit Care Med. 2013;188(6):733–748. doi: 10.1164/rccm.201308-1483ST.
- 11. Oka S, Furukawa H, Shimada K, et al. Serum biomarker analysis of collagen disease patients with acuteonset diffuse interstitial lung disease. *BMC Immunol.* 2013;14:9. doi: 10.1186/1471-2172-14-9.
- Travis WD, Hunninghake G, King TE Jr, et al. Idiopathic nonspecific interstitial pneumonia: report of an American Thoracic Society project. Am J Respir Crit Care Med. 2008;177(12):1338–1347. doi: 10.1164/ rccm.200611-1685OC.
- 13. Jeong YJ, Kim KI, Seo IJ, et al. Eosinophilic lung diseases: a clinical, radiologic, and pathologic overview. *Radiographics*. 2007;27(3):617–637. doi: 10.1148/rg.273065051.
- 14. Lacasse Y, Selman M, Costabel U, et al. Clinical diagnosis of hypersensitivity pneumonitis. Am J Respir Crit Care Med. 2003;168(8):952–958. doi: 10.1164/rccm.200301-137OC.
- Hunninghake GW, Costabel U, Ando M, et al. ATS/ERS/WASOG statement on sarcoidosis. American Thoracic Society/ European Respiratory Society/World Association of Sarcoidosis and other Granulomatous Disorders. Sarcoidosis Vasc Diffuse Lung Dis. 1999;16(2):149–173.
- Umehara H, Okazaki K, Masaki Y, et al. Comprehensive diagnostic criteria for IgG4-related disease (IgG4-RD), 2011. *Mod Rheumatol*. 2012;22(1):21–30. doi: 10.1007/s10165-011-0571-z.
- 17. Pérez-Dórame R, Mejía M, Mateos-Toledo H, et al. Rheumatoid arthritis-associated interstitial lung disease: lung inflammation evaluated with high resolution computed tomography scan is correlated to rheumatoid arthritis disease activity. *Reumatol Clin.* 2015;11(1):12–16. doi: 10.1016/j.reuma.2014.02.007.
- 18. Fu Y, Rong M, Zhu P, et al. The circulating fibrocytes are associated with the lung inflammation and fibrosis of mice with interstitial lung disease. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi.* 2014;30(8):814–818.
- 19. Fujiwara A, Kobayashi H, Masuya M, et al. Correlation between circulating fibrocytes, and activity and progression of interstitial lung diseases. *Respirology*. 2012;17(4):693–698. doi: 10.1111/j.1440-

1843.2012.02167.x.

- Hara Y, Shinkai M, Kanoh S, et al. Arterial carboxyhemoglobin measurement is useful for evaluating pulmonary inflammation in subjects with interstitial lung disease. *Intern Med.* 2017;56(6):621–626. doi: 10.2169/internalmedicine.56.7418.
- 21. Racanelli AC, Kikkers SA, Choi AMK, et al. Autophagy and inflammation in chronic respiratory disease. *Autophagy*. 2018;14(2):221–232. doi: 10.1080/15548627.2017.1389823.
- 22. Balestro E, Calabrese F, Turato G, et al. Immune inflammation and disease progression in idiopathic pulmonary fibrosis. *PLoS One*. 2016;11(5):e0154516. doi: 10.1371/journal.pone.0154516.
- 23. Kanoh S, Kobayashi H, Motoyoshi K. Exhaled ethane: an in vivo biomarker of lipid peroxidation in interstitial lung diseases. *Chest.* 2005;128(4):2387–2392. doi: 10.1378/chest.128.4.2387.
- 24. Faverio P, Bini F, Vaghi A, et al. Long-term macrolides in diffuse interstitial lung diseases. *Eur Respir Rev.* 2017;26(146):pii.170082. doi: 10.1183/16000617.0082-2017.
- Kinder BW, Brown KK, McCormack F, et al. Serum surfactant protein-A is a strong predictor of early mortality in idiopathic pulmonary fibrosis. *Chest.* 2009;135(6):1557–1563. doi: 10.1378/chest.08-2209.