INVITED REVIEW ARTICLE

Nagoya J. Med. Sci. 64. 103 ~ 108, 2001

MECHANISM-BASED THERAPY FOR LEUKEMIA: A LESSON FROM ATRA THERAPY

TOMOKI NAOE M.D., Ph.D.

Department of Infectious Diseases, Nagoya University Graduate School of Medicine

ABSTRACT

In the past two decades, there has been a tremendous increase in our understanding of the molecular mechanism of human leukemias. Leukemias are now recognized as a deregulated state of cell proliferation, differentiation and apoptosis, which is induced by gene alterations, including chromosomal translocations. Many of the mechanisms are potentially exploited as new targets for drug development. All-*trans* retinoic acid therapy for acute promyelocytic leukemia, which was initially developed as a differentiation therapy in an experienced-based manner, is currently known to be the first successful oncoprotein-directed therapy. Basic and clinical research into ATRA-resistance provides new directions for acute myeloid leukemia therapy will continue to lead the field of chemotherapy in the coming decades.

INTRODUCTION

Our comprehension of the molecular biology and pathophysiology of leukemia has advanced tremendously over past two decades. Non-random chromosomal translocations, found in nearly half of all leukemia cases, are closely linked to leukemogenesis.¹⁾⁻³⁾ Some of the oncogenes and anti-oncogenes found in solid tumors are also associated with leukemia.⁴⁾⁻⁶⁾ These gene alterations cause deregulated states of proliferation, differentiation and apoptosis, resulting in malignant hematopoiesis. Clinically, the molecular alterations are used as markers for diagnosis, detection of minimal residual disease and prediction of prognosis.^{7),8)}

On the other hand, therapeutic advances in leukemia have been independent of the above. Not only the chemotherapeutic concept, but also most anti-leukemia agents were established more than 20 years ago. Dose-escalation, modification of the schedule, and combinations of the agents were developed as treatments for leukemia.^{9),10)} In childhood acute lymphoblastic leukemia (ALL), over 80% patients are now cured.¹¹⁾ However, the prognosis of adult patients with acute leukemia has not significantly improved over the last decade.^{12),13)}

Notably, the combination of all-*trans* retinoic acid (ATRA) increased the complete remission rate and the number of long-term survivors.¹⁴⁾ The clinical use of ATRA for APL was first reported by the Shanghai group,¹⁵⁾ and was developed in an experience-based manner. The molecular studies later showed that ATRA directly targets the PML-RAR α oncoprotein generated by t(15;17) and modulates its function, resulting in differentiation and extinction of APL cells.¹⁶⁾ This clinical success emphasized the importance of developing new anti-leukemic therapy based on a different concept. Here, the author revisits ATRA therapy, and reviews the future directions of post-ATRA therapy.

Tsurumai-cho 65, Showa-ku, Nagoya 466-8560, Japan

TEL: 052-744-2955 FAX: 052-744-2801 E-mail: tnaoe@med.nagoya-u.ac.jp

PARADIGM SHIFT IN THE TREATMENT OF APL

In 1988, the Shanghai group reported the first clinical study of ATRA in 23 patients with APL. This treatment induced differentiation of blasts without bone marrow hypoplasia, followed by complete remission in 96% of the patients.¹⁵⁾ The effectiveness of ATRA therapy was confirmed in France¹⁷⁾ and Japan.¹⁴⁾ In the USA, a prospective randomized study showed a definitive advantage of ATRA-combined therapy.¹⁸⁾ However, it was unknown why ATRA was specifically effective against APL, although ATRA was known to be a non-specific inducer of differentiation. In 1990, two groups independently cloned the APL-specific chromosomal translocation t(15;17).^{19),20)} The translocation generates a chimeric gene between the PML gene on chromosome 15 and the retinoic acid receptor α (RAR α) gene on chromosome 17. It was soon believed that the resultant PML-RARa products influenced both the RARa and PML functions in a dominant negative manner.^{21),22)} Although the role of RAR α in myeloid differentiation remained unclear at this time, PML-RAR α was thought to block differentiation.¹⁶ Actually the above function of PML-RARα was confirmed by molecular studies in vitro and in vivo.²³⁾⁻²⁵ Then, the next question was why pharmacological concentrations of ATRA relieved the dominant negative function of PML-RARa. Since PML-RARa has a retinoic acid (RA)-binding region, ATRA was thought to directly bind to PML-RARa and modulate its function. Two important findings have accounted for the mechanism of modulation; First, immunohistochemical studies of PML showed that PML-RARa and PML are localized in diffuse microgranular patterns in the nucleus and cytoplasm.²⁶ This localization is restored to a nuclear microspeckled pattern by ATRA, which is caused by the degradation of PML-RAR α .²⁷⁾ Second, biochemical studies indicated that PML-RAR recruits a co-repressor complex including histone deacetylases (HDACs), which represses ATRA-dependent transcription.^{28),29)} At pharmacological concentrations of ATRA, PML-RARa binds the co-activator complex including CBP/p300 instead of the co-repressor complex, and undergoes transcription.^{28),29)} These two mechanisms seem to cooperatively relieve the dominant negative character of PML-RARa.

RETINOIDS-RESISTANCE IN APL

As the clinical usefulness of ATRA-therapy was confirmed, limitations of the sole or second ATRA-therapy became evident. If APL is treated by ATRA alone, the treated APL easily gains resistance to ATRA.³⁰⁾ The mechanism of the ATRA-resistance in vivo remains to be clarified. The major reason is reportedly the altered pharmaco-kinetics of ATRA; decreased absorption from the gastro-intestinal tract, sequestration in liver or other organs due to induction of cellular retinoic acid-binding protein-II, and rapid oxidation of ATRA by the p450 system.³⁰⁾ Notably, mutations of PML-RARa transcripts within its ligand-binding domain have been reported in relapsed APL,^{31),32)} although the cases are infrequent. Similar mutations have been also found in ATRA-resistant APL cell lines established in vitro.^{33),34)} Several different reported point mutations are localized in the E-domain, a ligand-binding domain, which is similar to ATRA-resistant HL 60 cells. Importantly, these mutations cluster at two particular regions within the E-domain. The crystal structure model of the RAR family suggests that these two cluster regions surround the ligand in the ligand-binding form. Studies of ATRA-resistance clarified that ATRA directly targets PML-RARa. Importantly, it should be noted that the mechanism of resistance in moleculetargeted therapy might be different from that in conventional chemotherapy, represented by multi-drug resistance.

TREATMENT OF ATRA-RESISTANT APL

ATRA-resistant APL has stimulated research for additional therapies. One approach is arsenic trioxide (As_2O_3) . Clinical use of As_2O_3 for APL began in North Eastern China in 1971,³⁵⁾ and was introduced to the world in 1997. *In vitro* experiments support the clinical efficacy of As_2O_3 .³⁶⁾ That is, treatment with 1 μ M As_2O_3 , a concentration that is clinically achievable, induces apoptosis in an APL cell line.³⁷⁾ A lower concentration of As_2O_3 causes morphological and immunophenotypic changes, although it does not induce terminal differentiation.³⁸⁾

Since As_2O_3 has been proven effective against APL with t(15;17) *in vivo*, PML-RAR α was first speculated to be associated with the sensitivity to As_2O_3 . Furthermore, PML-RAR α is down-regulated by As_2O_3 more rapidly than by ATRA.³⁷⁾ However, degradation of PML-RAR α and changes in PML-subcellular localization were similarly induced by As_2O_3 in As_2O_3 -sensitive and -resistant APL cell lines, suggesting that their contribution to apoptosis is small.³⁹⁾ As_2O_3 -treatment activated caspase 8 in a CD95-independent manner, but reduced glutathione concentration-dependently, which is different from the ATRA pathway.³⁹⁾

Another approach is a histone deacetylase inhibitor. The acetylated and deacetylated histones are regarded as the key machinery of transcriptional activation and repression, respectively.⁴⁰⁾ The p300/CBP and other coactivators have histone acetyltransferase activity associated with transcription.⁴¹⁾ Many investigators have also shown that histone deacetylases (HDACs) interact with inactive and/or non-liganded transcription factors via co-repressors such as mSin3A, N-CoR and SMRT to repress transcription in mammalian cells.^{42),43)} On the basis of molecular background, HDAC inhibitors (HDACI) such as butyrate, trichostatin A (TSA) and trapoxin A (TPX) were shown to block the repression, resulting in transcription of the target genes.⁴⁴⁾ Recently, three independent groups demonstrated *in vitro* that HDACI blocked the repression by PML-RARα. The combined therapy using ATRA and HDACI is effective in inducing differentiation in ATRA-resistant APL cells and cell lines.^{28),29)} Thus HDACI is considered as a promising agent for "differentiation therapy" in APL.

CHIMERIC TRANSCRIPTIONAL FACTORS

Differentiation therapy is theoretically applicable to all types of AML, because differentiationblock is one of the most important pathophysiological events in AML. Recent studies showed that chimeric transcriptional factors, generated by chromosomal translocations, are frequently associated with a differentiation-block. For example, in t(8;21), the fusion protein AML1-ETO recruits a co-repressor/HDAC complex. HDACs are also responsible for transformation by AML1-ETO, suggesting that HDAC is a common target for myeloid leukemias.^{42),43)} Strikingly, AML1-ETO expression blocks retinoic acid (RA) signaling in myelopoiesis.⁴⁵⁾ Accordingly, activation of the RA signaling pathway and inhibition of HDAC activity might represent a general strategy for differentiation induction in AML.

FUTURE DIRECTIONS

In the development of mechanism-based therapy, one of the most important issues is how to predict clinical efficacy. So far, leukemia cell lines have been used for screening chemotherapeutic agents. However, there are significant differences between *in vitro* and *in vivo* activities Transplanted or genetically modified mouse models of human leukemia are important not only for elucidating the mechanism of leukemia, but also for evaluating the *in vivo* efficacy.

After the success of ATRA, encouraging results emerged in the treatment of chronic myeloid leukemia. The effectiveness of a tyrosine kinase inhibitor (STI571), developed to inhibit abl kinase, was seen on clinical studies.⁴⁶⁾ This is the first example of a medicine developed in a mechanism-based and molecule-directed manner. The author believes that anti-leukemia therapy, which has led the field of chemotherapy from the middle of the 20th century onwards, will continue to be at the forefront of advances in cancer treatment.

REFERENCES

- Mrozek K., Heinonen K., de la Chapelle A. and Bloomfield CD.: Clinical significance of cytogenetics in acute myeloid leukemia. *Semin. Oncol.* 24: 17–31 (1997).
- Caligiuri MA., Strout MP. and Gilliland DG.: Molecular biology of acute myeloid leukemia. Semin. Oncol. 24: 32–44 (1997).
- 3) Look AT.: Oncogenic transcription factors in the human acute leukemias. Science. 278: 1059–1064 (1997).
- Neubauer A., Dodge RK., George SL., Davey FR., Silver RT., Schiffer CA., Mayer RJ., Ball ED., Wurster-Hill D., Bloomfield CD. and Liu ET.: Prognostic importance of mutations in the *ras* proto-oncogenes in de novo acute myeloid leukemia. *Blood.* 83: 1603–1611 (1994).
- 5) Nakano Y., Naoe T., Kiyoi H., Kitamura K., Minami S., Miyawaki S., Asou N., Kuriyama K., Kusumoto S., Shimazaki C., Akiyama H., Saito K., Nishimura M., Motoji T., Shinagawa K., Saito H. and Ohno R.: Prognostic value of p53 gene mutations and the product expression in de novo acute myeloid leukemia. *Eur J Haematol.* 65: 23–31 (2000).
- 6) Kiyoi H., Naoe T., Nakano Y., Yokota S., Minami S., Miyawaki S., Asou N., Kuriyama K., Jinnai I., Shimazaki C., Akiyama H., Saito K., Oh H., Motoji T., Omoto E., Saito H., Ohno R. and Ueda R.: Prognostic implication of FLT3 and N-RAS gene mutations in acute myeloid leukemia. *Blood.* 93: 3074–3080 (1999).
- 7) Willman CL.: Molecular evaluation of acute myeloid leukemias. Semin. Hematol. 36: 390-400 (1999).
- Macintyre EA., and Delabesse E.: Molecular approaches to the diagnosis and evaluation of lymphoid malignancies. *Semin. Hematol.* 36: 373-389 (1999).
- 9) Bishop JF.: The treatment of adult acute myeloid leukemia. Semin. Oncol. 24: 57-69 (1997).
- Laport GF. and Larson RA.: Treatment of adult acute lymphoblastic leukemia. Semin. Oncol. 24: 70–82 (1997).
- 11) Pui CH, and Evans WE.: Acute lymphoblastic leukemia. N. Engl. J. Med. 339: 605-615 (1998).
- 12) Baudard M., Beauchamp-Nicoud A., Delmer A., Rio B., Blanc C., Zittoun R. and Marie JP.: Has the prognosis of adult patients with acute myeloid leukemia improved over years? A single institution experience of 784 consecutive patients over a 16-year period. *Leukemia*. 13: 1481–1490 (1999).
- 13) Ohno R.: How high can we increase complete remission rate in adult acute myeloid leukemia? *Int. J. Hematol.* 72: 272–9 (2000).
- 14) Kanamaru A., Takemoto Y., Tanimoto M., Murakami H., Asou N., Kobayashi T., Kuriyama K., Ohmoto E., Sakamaki H., Tsubaki K., Yamada O., Oh H., Saito K., Matsuda S., Minato K., Ueda T. and Ohno R.: Alltrans retinoic acid for the treatment of newly diagnosed acute promyelocytic leukemia. Japan Adult Leukemia Study Group. *Blood.* 85: 1202–1206 (1995).
- 15) ME Huang., YC Ye., SR Chen., JR Chai., JX Lu., L Zhoa., LJ Gu. and ZY Wang.: Use of all-trans retinoic acid in the treatment of acute promyelocytic leukemia. *Blood.* 72: 567–572 (1988).
- 16) Grignani F., Fagioli M., Alcalay M., Longo L., Pandolfi PP., Donti E., Biondi A., Lo Coco F., Grignani F. and Pelicci PG.: Acute promyelocytic leukemia: from genetics to treatment. *Blood.* 83: 10–25 (1994).
- 17) Castaigne S., Chomienne C., Daniel MT., Ballerini P., Berger R., Fenaux P. and Degos L.: All-trans retinoic acid as a differentiation therapy for acute promyelocytic leukemia. I. Clinical results. *Blood.* 76: 1704–1709 (1990).
- 18) Tallman M. S., Andersen J. W., Schiffer C. A., Appelbaum F. R., Feusner J. H., Ogden A., Shepherd L., Willman C., Bloomfield C. D., Rowe J. M. and Wiernik P. H.: All-trans-Retinoic Acid in Acute Promyelocytic Leukemia. *N. Engl. J. Med.* 1997; 337: 1021–1028 (1997).
- Borrow J., Goddard AD., Sheer D. and Solomon E.: Molecular analysis of acute promyelocytic leukemia breakpoint cluster region on chromosome 17. *Science*. 249: 1577–1580 (1990).
- 20) de The H., Chomienne C., Lanotte M., Degos L. and Dejean A.: The t(15;17) translocation of acute promyelocytic leukaemia fuses the retinoic acid receptor alpha gene to a novel transcribed locus. *Nature*.

347: 558–561 (1990).

- 21) Kakizuka A., Miller WH Jr., Umesono K., Warrell RP Jr., Frankel SR., Murty VV., Dmitrovsky E. and Evans RM.: Chromosomal translocation t(15;17) in human acute promyelocytic leukemia fuses RAR alpha with a novel putative transcription factor, PML. *Cell*. 66: 663–674 (1991).
- 22) de The H., Lavau C., Marchio A., Chomienne C., Degos L. and Dejean A.: The PML-RAR alpha fusion mRNA generated by the t(15;17) translocation in acute promyelocytic leukemia encodes a functionally altered RAR. *Cell.* 66: 675–684 (1991).
- 23) Grignani F., Ferrucci PF., Testa U., Talamo G., Fagioli M., Alcalay M., Mencarelli A., Grignani F., Peschle C., Nicoletti I. and Pelicci PG.: The acute promyelocytic leukemia-specific PML-RAR alpha fusion protein inhibits differentiation and promotes survival of myeloid precursor cells. *Cell.* 74: 423–431 (1993).
- 24) Li-Zhen He., Carla Tribioli., Roberta Rivi., Daniela Peruzzi., Pier Giuseppe Pelicci., Vera Soares., Giorgio Cattoretti. and Pier Paolo Pandolfi.: Acute leukemia with promyelocytic features in PML/RAR transgenic mice. *PNAS*. 94: 5302–5307 (1997).
- 25) Li-Zhen He., Fabien Guidez., Carla Tribioli., Daniela Peruzzi., Martin Ruthardt., Arthur Zelent. and Pier Paolo Pandolfi.: Distinct interactions of PML-RARalpha and PLZF-RARalpha with co-repressors determine differential responses to RA in APL. *Nature Genetics.* 18: 126 (1998).
- 26) Koken MH., Puvion-Dutilleul F., Guillemin MC., Viron A., Linares-Cruz G., Stuurman N., de Jong L., Szostecki C., Calvo F., Chomienne C., Degos L., Puvion E. and de The H.: The t(15;17) translocation alters a nuclear body in a retinoic acid-reversible fashion. *EMBO J.* 13: 1073–1083 (1994).
- 27) Yoshida H., Kitamura K., Tanaka K., Omura S., Miyazaki T., Hachiya T., Ohno R. and Naoe T.: Accelerated degradation of PML-retinoic acid receptor alpha (PML-RARA) oncoprotein by all-trans-retinoic acid in acute promyelocytic leukemia: possible role of the proteasome pathway. *Cancer Res.* 56: 2945–2948 (1996).
- 28) Lin RJ., Nagy L., Inoue S., Shao W., Miller WH Jr. and Evans RM.: Role of the histone deacetylase complex in acute promyelocytic leukaemia. *Nature*. 391: 811–814 (1998).
- 29) Grignani F., De Matteis S., Nervi C., Tomassoni L., Gelmetti V., Cioce M., Fanelli M., Ruthardt M., Ferrara FF., Zamir I., Seiser C., Grignani F., Lazar MA., Minucci S. and Pelicci PG.: Fusion proteins of the retinoic acid receptor-alpha recruit histone deacetylase in promyelocytic leukaemia. *Nature*. 391: 815–818 (1998).
- Warrell RP Jr.: Retinoid resistance in acute promyelocytic leukemia: new mechanisms, strategies, and implications. *Blood.* 82: 1949–1953 (1998).
- 31) Imaizumi M., Suzuki H., Yoshinari M., Sato A., Saito T., Sugawara A., Tsuchiya S., Hatae Y., Fujimoto T., Kakizuka A., Konno T. and Iinuma K.: Mutations in the E-domain of RAR portion of the PML/RAR chimeric gene may confer clinical resistance to all-trans retinoic acid in acute promyelocytic leukemia. *Blood.* 2: 374–82 (1998).
- 32) Wei Ding., Yun-Ping Li., Lucio M. Nobile., George Grills., Ines Carrera., Elisabeth Paietta., Martin S. Tallman., Peter H. Wiernik. and Robert E. Gallagher.: Leukemic Cellular Retinoic Acid Resistance and Missense Mutations in the PML-RAR Fusion Gene After Relapse of Acute Promyelocytic Leukemia From Treatment With All-*trans* Retinoic Acid and Intensive Chemotherapy. *Blood.* 92: 1172–1183 (1998).
- 33) Shao W., Benedetti L., Lamph WW., Nervi C. and Miller WH Jr.: A retinoid-resistant acute promyelocytic leukemia subclone expresses a dominant negative PML-RAR alpha mutation. *Blood.* 89: 4282–4289 (1997).
- 34) Kitamura K., Kiyoi H., Yoshida H., Saito H., Ohno R. and Naoe T.: Mutant AF-2 domain of PML-RARalpha in retinoic acid-resistant NB4 cells: differentiation induced by RA is triggered directly through PML-RARalpha and its down-regulation in acute promyelocytic leukemia. *Leukemia*. 11: 1950–1956 (1997).
- 35) Sun HD., Ma L., Hu XC. and Zhang TD.: Ai-Lin I treated 32 cases of acute promyelocytic leukemia. *Chin. J. Integrat. Chin & West Med.* 12: 170 (1992).
- 36) Zhi-Xiang Shen., Guo-Qiang Chen., Jian-Hua Ni., Xiu-Shong Li., Shu-Min Xiong., Qian-Yao Qiu., Jun Zhu., Wei Tang., Guan-Lin Sun., Kan-Qi Yang., Yu Chen., Li Zhou., Zhi-Wen Fang., Yan-Ting Wang., Jun Ma., Peng Zhang., Ting-Dong Zhang., Sai-Juan Chen., Zhu Chen. and Zhen-Yi Wang.: Use of Arsenic Trioxide (As₂O₃) in the Treatment of Acute Promyelocytic Leukemia (APL): II. Clinical Efficacy and Pharmacokinetics in Relapsed Patients. *Blood.* 89: 3354–3360 (1997).
- 37) Guo-Qiang Chen., Xue-Geng Shi., Wei Tang., Shu-Min Xiong., Jun Zhu., Xun Cai., Ze-Guang Han., Jian-Hua Ni., Gui-Ying Shi., Pei-Ming Jia., Meng-Min Liu., Kai-Li He., Chao Niu., Jun Ma., Peng Zhang., Ting-Dong Zhang., Pascale Paul., Tomoki Naoe., Kunio Kitamura., Wilson Miller., Samuel Waxman., Zhen-Yi Wang., Hugues de The., Sai-Juan Chen. and Zhu Chen.: Use of Arsenic Trioxide (As₂O₃) in the Treatment of Acute Promyelocytic Leukemia (APL): I.As ₂O₃ Exerts Dose-Dependent Dual Effects on APL Cells. *Blood.* 89: 3345–3353 (1997).
- 38) Cai X., Shen Y-L., Zhu Q., Jia P-M., Yu Y., Zhou L., Huang Y., Zhang J-W., Xiong S-M., Chen S-J., Wang Z-Y., Chen Z. and Chen G-Q.: Arsenic trioxide-induced apoptosis and differentiation are associated respectively with mitochondrial transmembrane potential collapse and retinoic acid signaling pathways in acute

Tomoki Naoe

promyelocytic leukemia. Leukemia. 262-270 (2000).

- 39) Kitamura K., Minami Y., Yamamoto K., Akao Y., Kiyoi H., Saito H. and Naoe T.: Involvement of CD95-independent caspase 8 activation in arsenic trioxide-induced apoptosis. *Leukemia*. 14: 1743–1750 (2000).
- 40) Westin S., Kurokawa R., Nolte RT., Wisely GB., McInerney EM., Rose DW., Milburn MV., Rosenfeld MG. and Glass CK.: Interactions controlling the assembly of nuclear-receptor heterodimers and co-activators. *Nature*. 395: 199–202 (1998).
- Chakravarti D., LaMorte VJ., Nelson MC., Nakajima T., Schulman IG., Juguilon H., Montminy M. and Evans RM.: Role of CBP/P300 in nuclear receptor signalling. *Nature*. 383: 99–103 (1996).
- 42) Lutterbach B., Westendorf JJ., Linggi B., Patten A., Moniwa M., Davie JR., Huynh KD., Bardwell VJ., Lavinsky RM., Rosenfeld MG., Glass C., Seto E. and Hiebert SW.: ETO, a target of t(8;21) in acute leukemia, interacts with the N-CoR and mSin3 corepressors. *Mol Cell Biol.* 18: 7176–7184 (1998).
- 43) Kitabayashi I., Yokoyama A., Shimizu K. and Ohki M.: Interaction and functional cooperation of the leukemia-associated factors AML1 and p300 in myeloid cell differentiation. *EMBO J.* 17: 2994–3004 (1998).
- 44) Naoe T.: Histone deacetylase inhibitor. Cellular Mollecular Medicine. 2: 331-336 (2001) (in Japanese).
- 45) Thomas Pabst., Beatrice U. Mueller., Nari Harakawa., Claudia Schoch., Torsten Haferlach., Gerhard Behre., Wolfgang Hiddemann., Dong-ER Zhang. and Daniel G. Tenen.: AML1-ETO downregulates the granulocytic differentiation factor C/EBPalpha in t(8;21) myeloid leukemia. *Nature Medicine*. 7: 444–451 (2001).
- 46) Druker B. J., Talpaz M., Resta D. J., Peng B., Buchdunger E., Ford J. M., Lydon N. B., Kantarjian H., Capdeville R., Ohno-Jones S. and Sawyers C. L.: Efficacy and Safety of a Specific Inhibitor of the BCR-ABL Tyrosine Kinase in Chronic Myeloid Leukemia. *N Engl J Med.* 344: 1031–1037 (2001).